2022 H2 Paper 2

Section A

Answer all questions

- 1(a) Outline the main features of the cell theory. [3]
 - 1. The cell is the smallest unit of life ;
 - 2. All known living organisms are made up of cells ;
 - 3. All cells come from pre-existing cells ;
- (b) (i) Complete Table 1.1 to show the name and function of the structures labelled **A**, **B**, **C** and **D** in Fig. 1.1. [4]

label	name	function				
Α	mitochondrion	Site of <i>aerobic respiration</i> * where				
		<u>ATP* synthesis occurs;</u>				
В	nucleus	Contains DNA where <i>transcription*</i> occurs to produce <i>mRNA*</i> ;				
С	<u>Golgi*</u> body/apparatus	Modify lipids and proteins by glycosylation* to form glycolipids and glycolipids respectively; To sort and package proteins into different vesicles and target the proteins to different parts of the cell or for secretion out of cell;				
D	Rough endoplamic reticulum	Translation* of mRNA to protein/ protein synthesis by bound <u>ribosomes</u> *; <u>Glycosylation</u> * of proteins; <u>Transportation</u> of proteins in transport vesicles to the Golgi apparatus;				

- (ii) Compare the process of asexual reproduction in yeast with binary fission in bacteria. [2]
 - 1. Similarity both result in 2 genetically identical* daughter cells;
 - Difference budding involves the <u>evagination</u>* of the cell surface membrane to form a daughter cell but binary fission involves the <u>invagination</u>* of the cell surface membrane to separate the 2 daughter cells;

[Total: 9]

- 2 With reference to Fig. 2.1, describe the effect of increasing lopinavir concentration on the
- (a) inhibition of HIV protease. [3]
 - 1. As the <u>concentration of lopinvar increased</u>, the <u>percentage inhibition</u> increased;
 - 2. From <u>0.0 to 0.4 µmoldm⁻³ lopinavir concentration</u>, there is a <u>gradual increase</u> in <u>percentage inhibition</u>, from <u>0.0 to 8.0%</u>;
 - 3. From <u>0.4 to 1.6 µmoldm⁻³ lopinavir concentration</u>, there is a <u>steep increase</u> in <u>percentage inhibition</u> from <u>8.0 to 97.0%</u>;
 - 4. From 1.6 to 2.4 μmoldm⁻³ lopinavir concentration, the <u>pecentage inhibition starts to</u> <u>plateau</u> from about <u>97.0 to 99.0%;</u>
- (b) Explain how lopinavir inhibits HIV protease. [4]
 - 1. <u>Lopinavir</u> has a <u>3D conformation</u> that is <u>complementary in shape</u> and charge to the <u>active site* of HIV protease;</u>
 - 2. and lopinavir has a similar shape as the substrate/polyproteins;
 - Lopinavir <u>competes with the substrate/polyproteins</u> for the <u>active site</u> of HIV protease and <u>prevents the substrate from binding</u> to active site and hence <u>reduces the rate of</u> <u>reaction</u>;
 - 4. The binding of the inhibitor to the active site is not permanent/reversible;
 - 5. However, at <u>high substrate concentration</u>, the <u>effect of the inhibitor can be overcome</u> and <u>Vmax can be reached</u>;
- (c) Translation of viral RNA in cells infected with the human immunodeficiency virus (HIV) results in large polypeptides known as polyproteins. HIV protease cuts each of these viral polyproteins into smaller molecules.

Suggest how inhibition of HIV protease may limit the ability of HIV to reproduce. [3]

- 1. If HIV protease activity is inhibited, then <u>viral polyproteins cannot be cleaved;</u>
- 2. Hence proteins such as <u>reverse transcriptase</u>, <u>integrase</u>, <u>protease</u> and <u>capsid</u> <u>proteins</u> cannot be synthesised;
- 3. <u>Fully functional virions cannot be produced</u> and the ability for HIV to reproduce will be limited;

[Total: 10]

- 3 Name the structures labelled **P**, **Q** and **R** in Fig. 3.1. [3]
- (a)

P enhancer

- **Q** promoter
- **R** structural gene/ coding region

((b)	Com	pare	transcri	ption	in	prokar	votes	and	eukar	votes.	[4]	L
		•••••			P		P	,			,	L . 1	ł

Feature	Prokaryotes	Eukaryotes
Binding of RNA polymerase	RNA polymerase <u>binds directly</u> to the <u>promoter</u> * via the sigma factor;	RNA polymerase is <u>recruited by</u> <u>general transcription factors</u> * to bind to the <u>promoter</u> * forming the <u>transcription</u> initiation complex*:
Genes transcribed	Transcription of <u>several genes</u> <u>in an operon</u> controlled by one promoter;	Transcription of <u>one gene</u> controlled by one promoter;
mRNA formed	Polycistronic* mRNA is formed;	<u><i>Monocistronic</i></u> * mRNA is formed;
Upregulation of transcription	In <u>lac operon</u> , cAMP binds to <u>catabolic activator protein</u> * (CAP). <u>Active CAP</u> binds to <u>CAP binding site* in the</u> promoter region and increase affinity of RNA polymerase* binding to the promoter *, thus increasing the frequency of transcription;	<u>Activator</u> * binds to <u>enhancer</u> * causing bending of spacer DNA and <u>promoting the assembly of</u> <u>the transcription initiation</u> <u>complex</u> which will <u>increase</u> <u>frequency of transcription</u> ;
Downregulation of transcription	Binding of <u>repressor</u> * to <u>operator</u> * prevents the RNA polymerase binding to the promoter and hence preventing transcription. E.g. lac repressor bind to operator in lac operon/ trp repressor bind to operator in trp operon;	Binding of repressor * to silencer * will prevent the assembly of the transcription initiation complex which will decrease frequency of transcription;

- (c) Explain how the introduction of mRNA into the cytoplasm of adult cells can result in the production of iPSCs **and** why these iPSCs are footprint free. [4] How introduction of synthetic mRNAs can produce iPSCs:
 - 1. **Translation*** of the synthetic mRNAs will produce proteins which are needed to
 - 2. <u>switch off certain genes</u> in specialised adult cells <u>allowing them to de-differentiate</u>;
 - 3. and <u>switch on expression of certain specific stem cell genes</u> allowing the adult somatic cells to <u>achieve pluripotency;</u>

Why are these iPSCs footprint-free:

- 4. The <u>synthetic mRNAs do not enter the nucleus</u> and <u>will not intergrate with the genomic DNA</u> they <u>do not modify the genome</u> of the adult cells;
- 5. The <u>synthetic mRNAs are degraded after a while</u> and will not have a trace inside the cells;

[Total: 11]

- **4(a)** Explain how the end replication problem arises during DNA synthesis **and** describe the consequences of this problem. [5]
 - 1. Occurs during replication of linear DNA.
 - 2. Each round of <u>DNA replication</u> will result in the <u>shortening</u> of daughter molecules at <u>the **5**' ends of the telomeres</u>.
 - 3. because *RNA primers* at the 5'end* is removed, but **DNA polymerase** is <u>unable to</u> replace with DNA; (idea of end replication problem)
 - 4. result in overhangs at the <u>3' end of the telomeres</u>.
 - 5. However, since telomeres are non-coding, this ensures that <u>vital genetic</u> <u>information/genes are not lost / eroded</u> with each round of replication;
 - 6. If the telomeres shorten to critical length, cell will be signalled to stop dividing;
- (b) (i) With reference to Fig. 4.1, describe the role of the short piece of RNA that forms part of the telomerase complex. [3]
 - 5 nucleotides of the <u>telomerase RNA anneals</u> and forms <u>complementary base</u> <u>pairs</u>* with the <u>single-stranded overhang at 3' end</u> of the telomere;
 - 2. which aligns the telomerase reverse transcriptase with respect to the DNA;
 - 3. The telomerase RNA then serves as the <u>template</u>* for formation of a complementary DNA sequence;
 - whereby <u>adenine</u>* pairs with <u>uracil</u>*, <u>thymine</u>* with <u>adenine</u>*, <u>cytosine</u>* with <u>guanine</u>*, and <u>guanine</u>* with <u>cytosine</u>*;
 - 5. Resulting in tandem repeat sequences;
 - (ii) With reference to Fig. 4.1, describe the activity of the reverse transcriptase part of the telomerase complex. [3]
 - Telomerase has an <u>active site</u>* that is <u>complementary in conformation and</u> <u>charge</u>* to a <u>specific</u>* <u>telomeric DNA sequence</u>;
 - Using <u>telomerase RNA</u> as a <u>template</u>*, telomerase reverse transcriptase forms a complementary <u>DNA</u>* sequence through <u>complementary base pairing</u>*; (whereby adenine base pair with uracil, thymine with adenine, cytosine with guanine, and guanine with cytosine)
 - 3. Catalyzes the formation of *phosphodiester bonds** between deoxyribonuceotides;
 - 4. translocation in the $5' \rightarrow 3'$ direction to produce a series of tandem repeats of <u>GGTTAG</u>, thus elongating the telomere/DNA at the 3' overhang;

[Total: 11]

- **5(a)** (i) a loss of function mutation of the tumour suppressor gene *p*53. [2]
 - 1. loss-of-function mutation in <u>both alleles/two copies</u> of p53 results in a <u>non-functional gene product/gene product/p53 protein not produced;</u>
 - this results in the <u>inability to inhibit cell cycle</u> or <u>inability to repair damaged DNA</u> or <u>inability to promote **apoptosis***/**programmed cell death***; therefore cells with damaged DNA can continue to divide
 </u>
 - (ii) a gain in function mutation of the proto-oncogene ras. [2]
 - 1. <u>Gain-of-function mutation*</u> such as <u>proto-oncogenes*</u> to be converted to <u>oncogenes*</u>;
 - 2. will result in <u>overexpression of proteins/growth factors</u> or <u>production of</u> <u>hyperactive/degradation resistant proteins/growth factors</u> or <u>promote</u> <u>progression of cell cycle</u>; therefore cells will proliferate

- (b) Describe the process of angiogenesis and explain its importance in the development of cancer. [4]
 - 1. angiogenesis is the process of <u>forming new blood vessels</u> in <u>response to chemical</u> <u>signals</u> given off by the tumour;
 - 2. blood vessels are essential to <u>transport blood with oxygen and nutrients</u> to the tumour;
 - 3. which are necessary for <u>cell growth and cell division</u> and hence the potential unlimited growth of the tumour;
 - blood vessels also provide a <u>route</u> for tumour cells to <u>metastasise to other parts of</u> <u>the body to form secondary tumours</u>; resulting in a malignant cancer
- (c) Suggest and explain one reason why the recorded incidence of this cancer is rising. [2]
 - 1. <u>increased awareness and better diagnostic technology</u> for breast cancer;
 - 2. could have led to <u>more cases being detected</u> and therefore a <u>steady increase</u> in incidence detected from <u>25 cases per 100 000 women 1976 to 65 cases 2015;</u>

OR

- 3. <u>changes in lifestyle;</u>
- 4. like <u>poor diet and high stress levels working women</u> (due to more women entering the workforce) led to higher incidence in breast cancer from 1976 to 2015;

OR

- 5. <u>better healthcare</u> and treatments for cancer patients;
- 6. allowed more women with breast cancer to have children and <u>pass breast cancer</u> <u>causing alleles to their children;</u>

OR

- 7. women <u>bearing children at when they are older;</u>
- 8. increases likelihood of <u>accumulation of more mutations</u> in <u>cancer causing alleles</u> <u>within eggs;</u>

Must quote data

[Total: 10]

6(a) Draw a genetic diagram to show the expected phenotypic ratio of the F2 offspring, assuming normal dihybrid inheritance.

Use the symbols **A** and **a** to represent the alleles for grain texture and **B** and **b** to represent the alleles for grain colour. [5]

Let **A** be the allele to code for smooth grain texture and **a** the allele to code for wrinkled grain texture while **B** is the allele that codes for purple grains and **b** is the allele that codes for yellow grains.

Parental phenotype	Smooth,	purple grains	x	X Wrinkled, yellow grains				
Parental genotype	1	ABB	Х	aabb				
Gametes		AB		ab				
F ₁ phenotype	All smooth, purple grains							
genotype			AaBb					
Self- pollination of F ₁ generation	AaBb x AaBb							
	Gametes	AB	Ab	aB	ab			
	AB	AABB	AABb	AaBB	AaBb			
		smooth purple grain	smooth purple grain	smooth purple grain	smooth purple grain			
		AABb	AAbb	AaBb	Aabb			
F ₂ generation:	Ab	smooth purple grain	smooth yellow grain	smooth purple grain	smooth yellow grain			
		AaBB	AaBb	aaBB	aaBb			
	ав	smooth purple grain	smooth purple grain	wrinkled purple grain	wrinkled purple grain			
	ab	AaBb	Aabb	aaBb	aabb			
		smooth purple grain	smooth yellow grain	wrinkled purple grain	wrinkled yellow grain			

 F2
 9 A_B_:
 3 A_bb:
 3 aaB_:
 1 aabb

 genotypes:
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F ₂	9 smooth	3 smooth	3 wrinkled	1 wrinkled
phenotype:	purple grain	yellow grain	purple grain	yellow grain

1. Corresponding parental phenotypes and genotypes

- 2. All gametes drawn circled
- 3. Corresponding F₁ phenotypes and genotypes
- 4. Corresponding F₂ phenotypes and genotypes (in Punnett Square)
- 5. Correct phenotypic ratio
- (b) One of the ears of maize resulting from self-pollination of the F1 plants had 216 smooth purple grains, 65 smooth yellow grains, 79 wrinkled purple grains and 21 wrinkled yellow grains.

A chi-squared test was carried out on these results to compare the expected number of each phenotypes with the observed number.

The calculated chi-squared value was 1.73.

The critical chi-squared value for these results at a probability of p = 0.05 is 7.81.

State **and** explain what can be concluded about inheritance of grain colour in maize from the chi-squared value of 1.73. [3]

- 1. The inheritance of grain colour in maize follows <u>normal Mendelian monohybrid</u> <u>inheritance;</u>
- 2. Since the χ^2 = 1.73, it is less than the critical χ^2 value of 7.81,
- 3. and we do not reject the null hypothesis;
- 4. There is <u>no significant difference between the observed and expected phenotypes</u> in the offspring generation and any difference is <u>due to chance only</u>;
- (c) Suggest how pure-breeding maize with smooth yellow grains can be obtained from the F2 grains by selection and breeding. [3]
 - 1. Perform a cross between the smooth, yellow grain from the F₂ generation with **homozygous recessive*** wrinkled yellow grains;
 - 2. If <u>all the offspring</u> from the cross are <u>smooth</u>, <u>yellow grains</u>, then the <u>original parent</u> <u>grain was a pure-breeding smooth</u>, <u>yellow grain</u>;
 - 3. If the <u>offspring</u> from the cross <u>produced a ratio of 1 smooth yellow grain : 1 wrinkled</u> <u>yellow grain</u>, then the <u>original parent grain was not pure-breeding (i.e. Aabb)</u>;

[Total: 11]

7(a) Outline the role of each of the structures K, L, M and N in Fig. 7.1 in the conversion of light energy to chemical energy during the light-dependent reactions of photosynthesis.[6]

K – Photosystem II (PSII) <u>harvests light energy</u> from photons and <u>emit electrons</u> which is passed to L;

<u>Photolysis of water</u>* by water-splitting enzyme so that the <u>electron emitted can fill the</u> <u>electron hole in PSII;</u>

L – <u>Electron carrier and protein pump</u> which <u>uses energy loss from electron flow to pump</u> protons from *stroma**into the *thylakoid space**; Protons generate the proton gradient for chemiosmosis to occur;

M – Photosystem I <u>harvest light energy</u> from photons and emit electrons; Electrons are passed to <u>**NADP reductase**</u>^{*} and it catalyses the formation of <u>**NADPH**</u>^{*} from NADP;

N – <u>ATP synthase*</u> allows protons to pass through it from the *thylakoid space* into the *stroma** down a concentration gradient via *chemiosmosis**;

The <u>electron motive force</u>* generated by the flow of protons is used to catalyse the <u>formation of ATP</u>* from ADP and Pi;

- (b) Describe the role of reduced NADP in linking the light-dependent reactions to the Calvin cycle. [2]
 - 1. <u>NADPH</u> synthesized in the light dependent reaction is needed for the Calvin cycle to <u>occur</u>, hence they link the 2 reactions;
 - In the Calvin cycle, <u>glycerate phosphate*</u> is reduced to <u>glyceraldehyde-3-</u> <u>phosphate *(</u>3C). NADPH provides the <u>reducing power</u> for the reaction as high energy electrons are stored in it;
- (c) Explain why oxygen is not produced during cyclic photophosphorylation. [2]
 - In cyclic photophosphorylation, <u>energy from light</u> will be absorbed by PSI and <u>be</u> <u>channeled to P700 of PSI</u> which will be excited and displace an electron that will be <u>accepted by the primary electron acceptor Y</u> and subsequently transferred to the <u>middle of the first electron transport chain;</u>
 - 2. The <u>electron is then transported down the **electron transport chain**^{*} (ETC) and is finally <u>recycled back to PSI filling the electron hole</u> hence <u>there is no photolysis</u> <u>needed to emit electrons to fill the electron hole</u> and no oxygen produced;</u>

[Total: 10]

- 8(a) Name the structures labelled V, W, X, Y and Z in Fig. 8.1. [5]
 - V ligand
 - W G-protein coupled receptor
 - X adenylyl cyclase
 - Y cAMP
 - Z relay protein/ kinase protein (A: protein kinase A)

- (b) Outline the stages in cell signalling that occur **after** the initial stages shown in Fig. 8.1. [5]
 - 1. <u>Signal transduction is initiated</u> when activation of the intracellular domain of the GPCR, upon ligand binding, will enable the <u>inactive G protein nearby to bind</u> to it;
 - 2. <u>G-protein is activated</u> when is <u>displaces its attached GDP</u> for GTP;
 - The <u>activated (α subunit of the) G-protein</u> will <u>translocate along membrane</u> and <u>bind</u> to enzyme <u>adenylyl cyclase</u>* and phosphorylate it, thus activating it;
 - 4. Adenylyl cyclase will <u>catalyze conversion of **ATP***</u> to **cAMP***;
 - 5. Signal amplification begins from <u>cAMP</u>* where the <u>number of activated product is</u> <u>always greater</u> than those in the preceding step as one moves down the pathway;
 - 6. <u>4 cAMP bind to and activates the relay protein/kinase protein/ protein kinase A (PKA)</u> causing it to detach into subunits;
 - 7. The <u>two activated subunits</u> of the relay protein/kinase protein/ <u>protein kinase A (PKA)</u> will <u>initiate a sequential activation of kinases</u> resulting in a <u>phosphorylation cascade</u> <u>during which further amplification of the signal will occur;</u>
 - 8. Which leads to the activation of glycogen phosphorylase which catalyses the breakdown of glycogen to glucose (i.e. glycogenolysis in liver & muscle cells)
 - 9. During the cellular response, there will also be a decrease in the rate of glycolysis, and an increase in the rate lipid and protein breakdown;

[Total: 10]

- **9(a)** With reference to Fig. 9.1, explain why the Sumatran, Tapanuli and Bornean orangutans are considered to be separate species. [2]
 - 1. The <u>Sumatran and Bornean orangutans</u> have a <u>larger difference in DNA</u> <u>characteristics for variable 2 compared to variable 1</u> while the <u>Sumatran and Tapanuli</u> <u>orangutans</u> have a <u>larger difference in DNA characteristics for variable 1 compared</u> <u>to variable 2 and the Bornean and Tapanuli have a larger difference in DNA</u> <u>characteristics for both variables 1 and 2</u>;
 - Due to the <u>large differences in DNA characteristics</u>, between all 3 species they are <u>likely to have evolved from a common ancestor</u> a very long time ago and <u>accumulated</u> <u>many mutations over time</u> and eventually formed different <u>reproductively isolated*</u> <u>species</u> that are <u>unable to interbreed to form fertile*</u>, <u>viable*</u> offspring;
- (b) Explain how these three species of orangutan may have evolved from a common ancestor. [3]
 - When <u>sea levels rose</u> at the <u>end of the ice age when ice melted</u>, the ancestors of the orangutans became <u>geographically isolated</u>* (A:a reproductive barrier formed). Thus the 3 sub-populations of orangutans were <u>prevented from interbreeding</u> and <u>gene flow was disrupted</u>*;
 - 2. <u>Different environments/niches</u> presented <u>different selection pressures</u>* and so individuals with <u>favourable traits</u> and were best adapted had a <u>selective advantage</u> and were <u>selected for</u>;
 - 3. and <u>survived and reproduced and passed on their favourable alleles</u> to their offspring which led to an <u>increasing frequency of favourable alleles</u>;
 - 4. As the 3 different sub populations <u>evolved independently of each other</u>, their <u>allele frequencies changed</u> as they <u>accumulated different genetic mutations</u>*, and were subjected to <u>genetic driff</u>* and <u>natural selection</u>*. Over a long period of time this led to <u>reproductive isolation</u>* and formation of 3 distinct but closely related orangutan species (i.e. macroevolution occurred);

- (c) Describe the advantages of using molecular methods to classify organisms. [3]
 - 1. Nucleotide data are <u>objective</u>. <u>Molecular character states are unambiguous</u> as A, C, G and T are easily recognisable and cannot be confused;
 - 2. Nucleotide data are <u>quantitative</u>. Molecular data are <u>easily converted to numerical</u> <u>form</u> and hence are amenable to mathematical and <u>statistical analysis</u> and hence computation. <u>Degree of relatedness can be inferred and quantified</u> by calculating nucleotide differences between species;
 - 3. Nucleotide data can be used to <u>compare species which are morphologically</u> <u>indistinguishable</u> especially if they are very closely related;
 - 4. As changes in nucleotide sequences accumulate over time with clockwork regularity. We can estimate the time of speciation of modern to ancient species;

[Total: 8]

- **10** Using the information provided, calculate the total number of different antigen-binding
- (a) sites for immunoglobulin molecules that are possible as a result of somatic recombination. [2]
 (Number of possible κ chain x number of possible heavy chains) + (Number of possible λ chain x number of possible heavy chains)
 = [(40 x 5) x (40 x 25 x 6)] + [(30 x 4) x (40 x 25 x 6)]
 = 1 920 000
- (b) Following infection or vaccination, there is a progressive increase in the affinity (binding ability) of antibodies for the antigen that has been introduced.

This progressive increase is called affinity maturation.

Explain how this process occurs. [3]

Somatic hypermutation / aka affinity maturation

- 1. Activated B cells divide rapidly by mitosis during clonal expansion*
- 2. some of these B cells can undergo somatic hypermutation, <u>mutation*</u> occurs on the <u>Ig genes Ig / gene that code for antibodies</u>.
- 3. each of these B cells will acquire slight <u>amino acid difference in the **variable regions** <u>of Ig chains / antigen-binding sites of BCR</u> on cell surface membrane).</u>
- The <u>B cells</u> that contains <u>BCR with higher affinity</u> for the antigen will go be selected to go through further <u>clonal expansion*</u> and <u>differentiation*</u> to form <u>plasma cells</u>*.

[Total: 5]

- 11 Explain how increases in temperature and rainfall could lead to an increase in the number
- (a) of cases of viral dengue disease. [2]
- Increase in temperature
 - 1. would lead to <u>increased rate of enzyme catalysed reactions</u> in the <u>mosquito</u> <u>vector Aedes aegypti</u>;
 - 2. leading to <u>faster development rate</u> and <u>decreased length of reproductive cycles</u> and mosquito <u>population increase</u> and hence greater spread of viral dengue disease;

Increase in rainfall

- 1. may lead to more pools of stagnant water;
- 2. which <u>female mosquitoes lay eggs</u> in and for the larval and pupal stages of the mosquito life cycle hence greater spread of viral dengue disease;

- (b) With reference to Fig. 11.1, evaluate how effective the scientists' model is in predicting the number of cases of viral dengue disease. [3] Effective
 - 1. the model <u>accurately predicted a peak exceeding the epidemic threshold of 192</u> <u>cases of viral dengue disease</u> from <u>week 26 to week 32</u> as both the predicted and reported data exceeded the threshold in the same weeks;
 - The model accurately <u>predicted the overall trend</u> of <u>low number of cases of</u> <u>around 80 cases</u> of viral dengue disease from <u>week 0 to week 10</u> and <u>sharp</u> <u>increase</u> from <u>week 10 to around week 31</u> (A: sharp decrease with relevant data quoted);

Not entirely effective

- 3. The actual peak <u>exceeded the predicted peak</u> of 216 cases by <u>50 cases</u> (actual peak was 266 cases) indicating the spread was more severe than predicted;
- 4. There were <u>periods of inaccurate predictions</u> e.g. predicted a <u>peak</u> in <u>week 3</u>, instead there was a <u>minimum number</u> of cases in <u>week 5</u>;

[Total: 5]

- End of Paper -